

Original Research Article

<https://doi.org/10.20546/ijcmas.2020.906.058>

Study on Optimization of Must and Fermentation Conditions for Production of Jack Fruit (*Artocarpus heterophyllus* L.) Wine

S. K. Anil¹, Praveen Gidagiri^{2*} and Praveen Jholgikar³

¹Department of Post-Harvest Technology, College of Horticulture, Arabhavi, India

²Department of Post-Harvest Technology, College of Horticulture, Bagalkot, India

³Department of Fruit Science, College of Horticulture, Arabhavi, India

*Corresponding author

ABSTRACT

The present investigation on Optimization of *must* and fermentation conditions for production of jack fruit (*Artocarpus heterophyllus* L.) wine was carried out in the department of Post-Harvest Technology, Kittur Rani Channamma College of Horticulture (University of Horticultural Sciences, Bagalkot), Arabhavi, during the year 2010-2011. It consisting of different treatments viz T₁- *Must* with 24°Brix TSS + 3.2 pH+1:2 dilution, T₂- *Must* with 24°Brix TSS+ 3.3 pH+1:2 dilution, T₃- *Must* with 24°Brix TSS + 3.4 pH+1:2 dilution, T₄- *Must* with 24°Brix TSS + 3.5 pH+1:2 dilution, T₅- *Must* with 24°Brix TSS + 3.2 pH+1:2.5 dilution, T₆- *Must* with 24°Brix TSS + 3.3 pH+1:2.5 dilution, T₇- *Must* with 24°Brix TSS + 3.4 pH+1:2.5 dilution and T₈- *Must* with 24°Brix TSS + 3.5 pH+1:2.5 dilution. The experiment was laid out in a completely randomized design with three replications. The main objective was to standardize the optimum level of dilution and pH in the jack fruit must. Various biochemical and organoleptic quality of wine recorded at regular interval during the storage of wine. The TSS, total sugars, reducing and non-reducing sugar content in all the treatments decreased following fermentation. The least TSS and sugars were observed in the wine made from *must* with 1:2.5 dilution + 3.4 pH (T₇) in the fresh wine as well in the aged wine under cold condition and in *must* with 1:2.5 dilution + 3.5 pH (T₈) under ambient condition. The left over sugars in the fresh wine had a range from 8.86 (T₈) to 10.79 (T₄) per cent respectively. Tannins per cent were showed non-significant difference and decreasing trend can be seemed over period of aging, T₈ (*must* with 1:2.5 dilution + 3.5 pH) observe the highest per cent of tannins throughout the investigation. The clarity of wine as revealed by maximum transmission (1.56 and 1.67) was noticed in T₈ and T₃ aged under cold and ambient condition.

Keywords

Post-Harvest Technology dilution 24°Brix TSS sugar content

Article Info

Accepted:

15 May 2020

Available Online:

10 June 2020

Introduction

The fruit is said to grow on the branches, the trunk, and the roots of the tree and looks like stuffed tripe hung all over the tree Jackfruit is quite popular in Eastern and Southern India and is cultivated widely in many states. The total area under jackfruit in India is approximately 1.02 lakh hectares. It is cultivated in an area of about 0.11 lakh hectares mostly in Southern Plains and

Western Ghats of Karnataka producing about 2.6 lakh tonnes of fruits per annum (Anon., 2000a). Jackfruit is not easy to eat out of hand owing to difficulty in separating fruit bulbs from rind.

This difficulty has its origin in morphological and biochemical hindrances associated with the fruit. Further the fruit in fresh form are not liked by many due to its intense flavour (Bhatia *et al.*, 1956). Therefore, it is urged

that indigenous or underutilized fruits which are not easily marketed in the fresh form should be processed into acceptable products (Jagadeesh *et al.*, 2007). The jackfruit is classified into two groups, one with firm to crisp flesh; generally highly flavoured, while other is generally mushy or soft fleshed (Singh, 1995). Jackfruit is nutritious, rich in vitamins (A and B), minerals (Ca, K and Fe) and contains considerable amounts of carotene and vitamin C. It is a rich source of pectin and protein (Nandini *et al.*, 1989).

Jackfruit is a poor man's fruit which is available in plenty during the season (April-August) and firm flesh bulbs are marketed and relished as dessert fruit. However, the soft flesh type does not find utility in the ripe form as it is not liked due to mashy texture. More than 60-70 per cent of the soft flesh form is wasted without utilization in the Western Ghats area where jackfruit is grown in plenty. This form being rich in sugars can be suitably used for the production of fermented beverage like wine. This endeavour helps not only to utilize the underutilized soft flesh jackfruit type but also to exploit excess produce of the jackfruit during the season. It also ensures the development of a sustained jackfruit processing cottage industry in rural areas and small towns.

Materials and Methods

Total soluble solids (°Brix)

The total soluble solids of the wine were determined with the help of "ERMA" hand refractometer having a range of 0 to 32 °Brix at room temperature.

pH

The pH of the wine was measured using digital pH meter (Model: Analog, research, USA). Standard buffer solution of pH 4.01

and 9.08 were used as reference to calibrate the instrument.

Titrateable acidity (%)

Ten ml of sample was taken in a 100 ml volumetric flask and the volume was made up. From this 10 ml of aliquot was taken in a 100 ml conical flask and titrated against 0.1 N NaOH using one or two drops of Phenolphthalein indicator. Appearance of light pink color denoted the end point. Total titrateable activity was expressed as per cent citric acid. (Srivastava and Kumar, 1993)

% of acidity =

$$\frac{\text{Titrateable value} \times \text{Normality of NaOH} \times \text{Vol. made up} \times \text{Equivalent wt. of acid}}{\text{Vol. of sample for estimation} \times \text{Weight or volume of sample taken} \times 1000}$$

Estimation of ethanol

Determination of ethanol concentration in aqueous solutions by using alcohol meter. Alcohol meter was directly dipped in the sample taken in a 500ml measuring cylinder and the scale in the meter where it coincides with the meniscus of the sample indicates per cent v/v alcohol present in the sample.

Estimation of tannins

Tannins were estimated by determining their oxidisability of potassium permanganate solution. A known volume of aliquot was taken in a porcelain dish. Indigo carmine solution (20 ml) and water (about 500-750 ml) were added and titrated against potassium permanganate solution until the colour turns to bright yellow or to a faint pink at the rim. Milliliters of potassium permanganate used (A) was noted.

A known volume of sample was taken and added with 25 ml of gelatin solution and the volume was made up to 500 ml by adding acid sodium chloride solution. From that, a 50

ml sample was taken and added with 20 ml of indigo carmine solution. Again the volume was made up to 500 ml by adding water and titrated against potassium permanganate solution as before (B).

The amount of milliliters of $KMnO_4$ used in this case (B) subtracted from (A) gave the quantity of $KMnO_4$ solution required to oxidise the tannins. The percentage of tannins in the sample was calculated by the formula given below.

A: Total tannin like materials

B: Non- tannin materials

A– B: True tannins

% of tannin =

$$\frac{\text{Titration value} \times \text{g of tannins/ml of } KMnO_4}{\text{Vol. of sample for estimation}} \times 100$$

1 ml of 0.1 N $KMnO_4$ = 0.0042 g of tannin

Results and Discussion

TSS (°B)

In the present study, TSS of fresh jack fruit wine recorded a range from 9.00 to 10.93 °B from the initial 24 °B (Table 1). The treatment T_7 (*Must* with 1:2.5 dilution+3.4 pH) recorded significantly lowest TSS of 9.00°B. The highest TSS (10.93 °B) was found in treatment T_4 (*Must* with 1:2 dilution + 3.5 pH). The TSS content of wines decreased slightly in all the treatments during aging irrespective of the conditions of aging. Aging of resultant wines under cold condition ($14 \pm 1^\circ C$), resulted a significantly lower TSS (8.73°B) in the treatment T_7 (*Must* with 1:2.5 dilution+3.4 pH) followed by T_8 and T_6 (Table1). The highest TSS of 10.31 °B was found in treatment T_4 (*Must* with 1:2 dilution + 3.5 pH) at 3 months and in the treatment T_1 (9.73°B) at 6 months. The treatment T_7 continued to remain low in terms of TSS at

the end of 6 months of aging. Aging under ambient condition ($27 \pm 1^\circ C$) resulted a lower TSS (8.71 and 8.26) in the treatment T_8 (*Must* with 1:2.5 dilution+3.5 pH). The highest TSS (10.06 °B) was found in treatment T_1 (*Must* with 1:2 dilution + 3.2 pH) at 3 months and 9.44 °B was found in T_2 at 6 months of aging. The reduction in the TSS during aging of wines is expected to be due to the slower yeast activity that may still prevail during aging which converts sugars into alcohol (Owen, 2004). A corresponding increase in alcohol content of respective treatments during aging corroborates the view that sugars are fermented by yeasts to produce a little more alcohol. The jack fruit wine witnessed further but slight decrease in the TSS during ageing both in cold and ambient condition ($14 \pm 1^\circ C$ and $27 \pm 1^\circ C$) with the same trend in all the treatments demonstrating slow activity of wine yeasts.

pH

In this study, pH of jack fruit *must* was adjusted to 3.2 to 3.5 with 2 different combinations of dilution levels. The pH of fresh jack fruit wine showed a slight decrease in all the treatments. The decrease in pH of wine in all treatments might be due to production of large amounts of acidic wastes. Wines vary considerably in pH, with values below 3.1 being perceived as sour and those above 3.7 being perceived as flat (Jackson, 1994). Only fresh wines in the treatments T_4 , T_6 and T_8 recorded the pH slightly above 3.0 with remaining treatments documenting a pH below 3.0 (Table 2).

However, during ageing all the treatments have recorded an increase in the pH values. Similar observations of increase in pH after fermentation and during ageing have been recorded by Shankar *et al.*, (2004) in guava wine. After 6 months of ageing, jack fruit wine with the treatment T_4 (*must* with 1:2

dilution + 3.5 pH) continued to maintain the tendency to record maximum pH (4.09 both in cold and ambient ageing) while the minimum (3.51 and 3.79) was observed in T5 (*Must* with 1:2.5 dilution + 3.2 pH) and T₆ (*Must* with 1:2.5 dilution + 3.3 pH). The degradation of arginine, one of the major amino acids in grape *must* and wine, releases ammonia, which may help to raise the pH (Jackson, 1994). Shukla *et al.*, (1991) analysed the pH of the wine samples prepared from different varieties of jambalfruit and the pH of these wines ranged from 3.50 to 3.40. The increase in pH was due to reduction in acidity through precipitation of potassium tartarate salts from wine or due to enhanced synthesis of esters from ethyl alcohol and volatile acids. The changes in the pH were not correlated with the changes in total acidity because of the buffering capacity of the wines and the relative amount of various acids influencing the acidity (Shankar *et al.*, 2004).

Titration acidity (%)

It is important to have the perfect balance of sugar, acidity and pH in the *must* for quality wine production. Titration acidity of wine depends upon the total organic acids content of the wine. The mean titration acidity content of the fresh jack fruit wine was 1.10 per cent while that after 3 and 6 months of ageing under cold condition was 1.07 and 1.02 per cent respectively (Table 3). The decrease in the acidity during ageing might be due to combination of acids with alcohol to form esters which adds aroma to the wine during ageing (Shankar *et al.*, 2004). The titration acidity is an important parameter used to measure the quality of wine (Olasupo and Obayori, 2003). The ideal acidity in the wine is dependent on the style and preferences of the consumer. Nevertheless, the acceptable range for total acidity in most wines is between 5.5 and 8.5 mg/l (0.55 to 0.85%). White wines are generally preferred at higher

end of the scale whereas red wines are more appreciated at lower end (Jackson, 1994).

After six months of ageing in cold condition jack fruit wine showed minimum titration acidity (0.81%) in T₈ (*must* with 1:2.5 dilution+ 3.5 pH) which was at par with T₄ (*must* with 1:2 dilution+ 3.5 pH) and T₆ (*must* with 1:2.5 dilution+ 3.3 pH). The maximum value for the parameter was observed in T₁ (1:2 dilution+ 3.2 pH) acidity level of 1.2 per cent which was on par with T₁ and T₂. After three and six months of ageing in ambient condition, the treatment T₈ (*must* with 1:2.5 dilution+ 3.5 pH) recorded the minimum acidity content (0.83 and 0.81%) and maximum acidity (1.28 and 1.26 %) was found in Treatment T₁ (1:2 dilution+ 3.2 pH). The variation in the titration acidity level due to treatment differences have been accounted in karonda wine (Bhajipale *et al.*, 1998), mahua wine (Yadav, 2009), mango wine (Thippesha *et al.*, 1997) and strawberry wine (Somesh *et al.*, 2009).

Ethanol (% V/V)

Ethanol is undisputedly the most important alcohol in wine. Ethanol is the principal organic by-product of fermentation (Jackson, 1994). Alcohol production is one of the parameters to test the efficiency of fermentation, since alcohol is a major solvent in wine. The highest ethanol content (8.85 %) in the fresh wine was noticed in treatment T₇ *Must* with 1:2.5 dilution + 3.4 pH (Table 4). It can be attributed to the higher amount of available sugars and the favourable *must* conditions (dilution and pH) for fermentation. During ageing, the alcohol level increased slightly in all the treatments. This increase might have been due to very slow fermentation that occurred during ageing. The T₇ (*must* with 24⁰Brix TSS + 3.5pH) treatment continued to exhibit significantly higher ethanol content (8.92, 9.01 and 9.09

%) after six months of ageing in cold and after three and six months of ageing in ambient condition respectively. Kotecha (2010a) recorded a similar range of alcohol content in wines of banana and ber (7.6%), jamun (8.4 %), sapota 7.2% and strawberry (7.0%). Somesh *et al.*, (2009) observed a variation in the alcohol content of strawberry wine; Reddy and Reddy (2009) in mango wine; and Joshi *et al.*, (2005) in peach wine due to variation in varieties. In some treatments, there was decrease in the alcohol content. Auto oxidation of alcohol to esters or aldehydes or due to combining ability of alcohol with acidity of wine to form esters was observed in guava wine (Shankar *et al.*, 2004).

Tannins (%)

Tannins are one of the important parameter in wine which gives subtle aroma and bouquet during ageing. Tannins non-significant differences among the treatments in fresh wine as well as in aged wines in all the periodical observations of ageing in both cold and ambient condition (Table 5). There was a decrease in the per cent of tannins in all the treatments, this may due to hydrolysis of poly phenols to isoprenoid (Jackson, 1994). Treatment T₈ (*Must* with 1:2.5 dilution + 3.5 pH) obtained the highest per cent of tannins whereas treatment T₆ (*Must* with 1:2.5 dilution + 3.3 pH) recorded the lowest tannin content throughout the investigation. The per cent of tannins may vary in the treatments depending upon the type of wine, yeast fermentation conditions, containers and the maturation period was observed in litchi wine (Singh and Kaur, 2009).

Wine recovery (%)

The wine recovery in this study showed significant variations due to the treatments differences. Higher wine recovery was noted

at higher (1:2.5) of the two dilutions employed in the present study (Table 6). Irrespective of the dilutions, an increase in the wine recovery was found associated with higher pH levels. The wine recovery recorded a wide range from 45.74 to 78.13 per cent which may be attributed to the difference in the dilution and pH of the *must*. The treatment T₈ (*Must* with 1:2.5 dilution + 3.5 pH) exhibited the highest recovery percentage of 78.13 followed by T₇ (67.72%) and T₆ (61.94%). The variation of wine recovery in this study may be attributed to the influence of pH and dilution level of *must* on the activity of yeasts. A combination of higher *must* dilution (1:2.5) and higher pH (3.5) was found to yield more wine. Bhajipale *et al.*, (1998) in karonda wine; and Saravana *et al.*, (2001) in strawberry reported differences in wine yield due to treatment differences.

Colour (OD value)

The colour of jack fruit wine as represented by OD values in fresh wine and after ageing in both ambient and cold conditions showed non-significant variation among the treatments. Higher OD values indicate lower transmission and in turn lower brightness and clarity. Table 7 shows the treatment T₈ (*Must* with 1:2.5 dilution + 3.5 pH) obtained higher OD value of 1.56 under cold aging conditions while T₃ (*Must* with 1:2 dilution + 3.4pH) after 6 months of ageing under ambient condition. The lowest OD value of 1.07 and 1.08 were associated with T₂ at 3 months of aging under both the conditions. However, at 6 months of aging the least OD of 1.07 and 0.98 was noticed in treatment T₆ (*Must* with 1:2.5 dilution + 3.3 pH) and T₁ (*Must* with 1:2 dilution + 3.2 pH) respectively under cold and ambient conditions. The greater wine clarity allows for more transmission of light resulting in smaller OD value whereas deeper colour of wine allows lesser transmission of light and increased OD value.

Table.1 Effect of ‘must dilution and pH’ on Total soluble solids (^oBrix) of fresh jack fruit wine and after ageing

Tr. No	Treatment details	TSS (^o Brix)					
		Must	Fresh wine	Ageing in cold		Ageing in ambient	
				3MA	6MA	3MA	6MA
T ₁	Must with 1:2 dilution + 3.2 pH		10.36	9.89	9.73	10.06	9.26
T ₂	Must with 1:2 dilution + 3.3 pH		10.13	9.70	9.33	10.03	9.44
T ₃	Must with 1:2 dilution + 3.4pH		10.46	9.73	9.13	9.86	9.20
T ₄	Must with 1:2 dilution + 3.5pH		10.93	10.32	9.26	9.76	8.80
T ₅	Must with 1:2.5 dilution + 3.2 pH		10.73	9.80	9.47	9.83	8.60
T ₆	Must with 1:2.5 dilution + 3.3 pH		9.30	9.20	9.07	9.20	8.76
T ₇	Must with 1:2.5 dilution + 3.4 pH		9.00	8.70	8.41	8.73	8.60
T ₈	Must with 1:2.5 dilution + 3.5 pH		9.03	9.00	8.67	8.71	8.26
	Mean	24	9.99	9.54	9.13	9.52	8.86
	S.Em ±	-	0.15	0.08	0.09	0.05	0.08
	CD 1%	-	0.48	0.27	0.29	0.19	0.26

3MA –3 months of ageing 6MA - 6 months of ageing
Ageing in cold at 14±1°C; Ageing in ambient condition 27±1°C

Table.2 Effect of ‘must dilution and pH’ on pH of fresh jack fruit wine and after ageing

Tr. No	Treatment details	pH				
		Fresh wine	Ageing in cold		Ageing in ambient	
			3MA	6MA	3MA	6MA
T ₁	Must with 1:2 dilution + 3.2 pH	2.54	3.62	3.95	3.56	3.81
T ₂	Must with 1:2 dilution + 3.3 pH	2.71	3.56	3.93	3.76	3.89
T ₃	Must with 1:2 dilution + 3.4pH	2.84	3.96	4.05	3.87	3.94
T ₄	Must with 1:2 dilution + 3.5pH	3.15	3.98	4.09	3.99	4.09
T ₅	Must with 1:2.5 dilution + 3.2 pH	2.82	3.46	3.51	3.68	3.81
T ₆	Must with 1:2.5 dilution + 3.3 pH	3.11	3.24	3.83	3.54	3.79
T ₇	Must with 1:2.5 dilution + 3.4 pH	2.81	3.63	4.02	3.88	4.04
T ₈	Must with 1:2.5 dilution + 3.5 pH	3.26	3.88	4.01	3.94	4.13
	Mean	2.91	3.66	3.92	3.77	3.93
	SEm ±	0.04	0.08	0.04	0.03	0.10
	CD 1%	0.14	0.26	0.14	0.12	0.21

Table.3 Effect of ‘must dilution’ and pH on Titrable acidity (%) of fresh jack fruit wine and after ageing

Tr. No	Treatment details	Titrable acidity (%)				
		Fresh wine	Ageing in cold		Ageing in ambient	
			3MA	6MA	3MA	6MA
T1	Must with 1:2 dilution + 3.2 pH	1.31	1.29	1.24	1.28	1.24
T2	Must with 1:2 dilution + 3.3 pH	1.19	1.15	1.12	1.06	1.12
T3	Must with 1:2 dilution + 3.4pH	1.08	1.06	1.01	1.07	1.01
T4	Must with 1:2 dilution + 3.5pH	1.01	0.98	0.91	0.99	0.91
T5	Must with 1:2.5 dilution + 3.2 pH	1.09	1.07	1.02	1.04	1.02
T6	Must with 1:2.5 dilution + 3.3 pH	0.98	0.94	0.89	0.97	0.89
T7	Must with 1:2.5 dilution + 3.4 pH	1.26	1.21	1.19	1.21	1.19
T8	Must with 1:2.5 dilution + 3.5 pH	0.87	0.85	0.81	0.83	0.81
	Mean	1.10	1.07	1.02	1.06	1.03
	SEm ±	0.05	0.03	0.04	0.06	0.04
	CD 1%	0.16	0.11	0.17	0.21	0.15

3MA –3 months of ageing 6MA - 6 months of ageing
Ageing in cold at 14±1°C; Ageing in ambient condition 27±1°C

Table.4 Effect of ‘must dilution and pH’ on Ethanol (%) of fresh jack fruit wine and after its ageing

Tr. No	Treatment details	Alcohol (%)				
		Fresh wine	Ageing in cold		Ageing in ambient	
			3MA	6MA	3MA	6MA
T₁	Must with 1:2 dilution + 3.2 pH	7.68	8.04	8.42	8.23	8.54
T₂	Must with 1:2 dilution + 3.3 pH	7.89	8.44	8.65	8.24	8.31
T₃	Must with 1:2 dilution + 3.4pH	7.69	8.13	8.77	8.34	8.39
T₄	Must with 1:2 dilution + 3.5pH	7.45	7.30	8.70	8.40	8.46
T₅	Must with 1:2.5 dilution + 3.2 pH	7.54	8.09	8.59	8.36	8.29
T₆	Must with 1:2.5 dilution + 3.3 pH	8.38	8.43	8.80	8.73	8.36
T₇	Must with 1:2.5 dilution + 3.4 pH	8.85	8.70	8.92	9.01	9.09
T₈	Must with 1:2.5 dilution + 3.5 pH	8.08	8.85	8.04	8.86	8.94
	Mean	7.94	8.24	8.61	8.52	8.54
	SEm ±	0.16	0.15	0.12	0.19	0.20
	CD 1%	0.52	0.49	0.41	0.59	0.63

3MA –3 months of ageing 6MA - 6 months of ageing
Ageing in cold at 14±1°C; Ageing in ambient condition 27±1°C

Table.5 Effect of ‘must dilution and pH on Tannins’ (%) of fresh jack fruit wine and after ageing

Tr. No	Treatment details	Tannins (%)				
		Fresh wine	Ageing in cold		Ageing in ambient	
			3MA	6MA	3MA	6MA
T ₁	Must with 1:2 dilution + 3.2 pH	0.10	0.09	0.09	0.10	0.09
T ₂	Must with 1:2 dilution + 3.3 pH	0.14	0.11	0.10	0.13	0.11
T ₃	Must with 1:2 dilution + 3.4pH	0.12	0.10	0.09	0.12	0.09
T ₄	Must with 1:2 dilution + 3.5pH	0.11	0.11	0.09	0.10	0.08
T ₅	Must with 1:2.5 dilution + 3.2 pH	0.09	0.07	0.07	0.09	0.09
T ₆	Must with 1:2.5 dilution + 3.3 pH	0.07	0.05	0.04	0.07	0.06
T ₇	Must with 1:2.5 dilution + 3.4 pH	0.17	0.16	0.12	0.17	0.14
T ₈	Must with 1:2.5 dilution + 3.5 pH	0.19	0.17	0.13	0.19	0.16
	Mean	0.99	0.86	0.73	0.97	0.82
	SEm ±	0.08	0.04	0.06	0.05	0.07
	CD 1%	NS	NS	NS	NS	NS

3MA –3 months of ageing 6MA - 6 months of ageing

Ageing in cold at 14±1°C; Ageing in ambient condition 27±1°C

Table.6 Effect of ‘must dilution and pH’ on wine recovery (%) of fresh jackfruit wine after fermentation

Tr. No	Treatment details	Wine recovery (%)
		Fresh wine
T ₁	Must with 1:2 dilution + 3.2 pH	50.743
T ₂	Must with 1:2 dilution + 3.3 pH	45.743
T ₃	Must with 1:2 dilution + 3.4pH	51.670
T ₄	Must with 1:2 dilution + 3.5pH	54.260
T ₅	Must with 1:2.5 dilution + 3.2 pH	57.960
T ₆	Must with 1:2.5 dilution + 3.3 pH	61.946
T ₇	Must with 1:2.5 dilution + 3.4 pH	67.720
T ₈	Must with 1:2.5 dilution + 3.5 pH	78.130
	Mean	58.52
	SEm ±	2.89
	CD 1%	7.09

Table.7 Effect of ‘must dilution and pH’ on colour(OD value) of fresh jack fruit wine and after ageing

Tr. No	Treatment details	Colour (OD value)				
		Fresh wine	Ageing in cold		Ageing in ambient	
			3MA	6MA	3MA	6MA
T1	Must with 1:2 dilution + 3.2 pH	1.00	1.08	1.33	1.05	0.98
T2	Must with 1:2 dilution + 3.3 pH	1.00	1.07	1.38	1.08	1.12
T3	Must with 1:2 dilution + 3.4pH	1.00	1.36	1.08	1.34	1.67
T4	Must with 1:2 dilution + 3.5pH	1.00	1.20	1.32	1.26	1.27
T5	Must with 1:2.5 dilution + 3.2 pH	1.20	1.39	1.38	1.28	1.34
T6	Must with 1:2.5 dilution + 3.3 pH	1.00	1.17	1.07	1.17	1.14
T7	Must with 1:2.5 dilution + 3.4 pH	1.00	1.18	1.50	1.23	1.02
T8	Must with 1:2.5 dilution + 3.5 pH	1.20	1.33	1.56	1.17	1.24
	Mean	1.10	1.33	1.23	1.20	1.23
	SEm ±	0.07	0.09	0.20	0.90	0.14
	CD 1%	NS	NS	NS	NS	NS

A slight but insignificant variation in wine colour observed in this study may have its roots in the biochemical interaction between the sugars, pH and other constituents of pulp, in addition to the effect of conditions on fermentation and ageing.

Joshi *et al.*, (2005) observed a variation in the colour intensity of wine made from peach cultivars where in the higher colour intensity was recorded in cv. Redhaven (OD of 0.19) and lower in cv. Stark Early Gaint (OD of 0.86). The change in colour of wine made from guavas at different stages of maturity was attributed to variation in the pigmentation of fruits (Anderson and Badrie, 2005).

Ageing process is primarily physico-chemical; higher temperature speeds and activates most of the reactions involved. For most wines, prolonged exposure to temperatures of 40°C or more results in deterioration in quality. Carbohydrates in the wine undergo Maillard and thermal degradation reactions turning the wine brown (Jackson, 1994). The two temperatures adapted in the present study did not reveal much colour change as even the

higher temperature was much below the deleterious temperature (40°C).

References

- Anderson, C. and Badrie, N. 2005, Physico-chemical quality and consumer acceptance of guava wines. *Haryana J. Hort. Sci.*,42(3): 223-226.
- Anonymous., 2000, *Horticultural Crop Statistics of Karnataka State at a Glance*, Government of Karnataka, Directorate of Horticulture, Lal Bagh, Bangalore., pp. 86-87.
- Bhajibale, B.J., Gupta D.N. and Mehta, 1998, Effect of different stages of ripening of fruit of karonda wine. *Indian Food Packer*, 54: 27-30.
- Bhatia B.S., Siddappa, G.S. and Giridhari L.L., 1956, Development of fruit product: Dried jack seeds and flour, roasted nuts and jack papad. *Indian Food Packer*, 10: 30.
- Bhatia, B.S. and Siddappa, G.S., 1955, Development production from jackfruit. Part I. *Indian Food Packer*, 9: 7.
- Jagadeesh, S.L., Kirankumar, G., Hegde, L.,

- Swamy, G.S.K., Reddy, B.S., Basavaraj, N. and Raghavan, S.V., 2007, Variability studies in physico-chemical parameters in Chips purpose jackfruit selection of hilly zone in Karnataka. 2007, *Karnataka J. Agric. Sci.*, 20(2): 346-349.
- Joshi, V.K., Sharma, S. And Bhushan, S. 2005, Effect of method of preparation and cultivar on the quality of strawberry wine. *Acta Aliment.*, 34(4): 339-355.
- Kotecha, P.M., 2010a, Composition, quality and legal requirements of wine, pp. 121-127. Compendium of winter school on value addition of grapes with special reference to wine making at Rahuri (*Maharashtra*), 5-25 feb 10, p. 1-198.
- Nandini, P.V. and Dommen, B.E., 1989, Quality parameters of selected mango cultivars. *J. Food Sci. Technol.*, 39: 379-383.
- Olasupo, N.A. and Obayori, O.S., 2003, Utilization of palm wine (*Elaeis guinensis*) for the improved production of Nigerian indigenous alcoholic drink Ogogoro. *J. Food Pro. Preserv.*, 27: 365-372.
- Owen, S.G., Majeed, M., Lawrence, A.W. and Linda, D.W., 2004, effects of pectolytic enzymes and anti-oxidants on the quality of dry wines made from pineapple (*Ananas comosus*. L. Merr) peel. *Food Agric, and Envi* ,2(2):135-142.
- Saravana, K., Manimegalai, R.G. and Hamaran, M., 2001, Wine preparation from strawberry. *Beverage and Food World.*, pp. 18-19.
- Shankar, S., Dilip, J. and Narayana, Y., 2004, Changes in chemical composition of guava wine during storage. *Indian Food Packer*, 12:56-58.
- Shukla, K.G., Joshi, M.C., Yadav, S. and Bisht, N.S., 1991, Jambal wine making: standardization of a methodology and screening of cultivars. *J. Food. Sci.Technol.*, 28: 142-144.
- Somesh, S., Joshi, V.K. and Ghanshyam, A., 2009, An overview on strawberry wine production technology, Composition, maturation and quality evaluation. *Natural Product Radianc*e, 8(4): 356-365.
- Srivastava, R.P. and Kumar, S., 1993, Important methods for analysis of fruits and vegetable and their products. *Fruit and Vegetable Preservation Principles and Practices*. 2nd ed. pp. 321-339.

How to cite this article:

Anil. S. K., Praveen Gidagiri and Praveen Jhologikar. 2020. Study on Optimization of Must and Fermentation Conditions for Production of Jack Fruit (*Artocarpus heterophyllus* L.) Wine. *Int.J.Curr.Microbiol.App.Sci*. 9(06): 438-447. doi: <https://doi.org/10.20546/ijcmas.2020.906.058>